

REVIEW

Open Access



Host-directed therapy for tuberculosis

Na Tian^{1†}, Hongqian Chu^{2†}, Qi Li¹, Hong Sun², Jingfang Zhang², Naihui Chu^{1*} and Zhaogang Sun^{2*}

Abstract

Current TB treatment regimens are hindered by drug resistance, numerous adverse effects, and long treatment durations, highlighting the need for 'me-better' treatment regimens. Host-directed therapy (HDT) has gained recognition as a promising approach in TB treatment. It allows the repurposing of existing drugs approved for other conditions and aims to enhance the effectiveness of existing anti-TB therapies, minimize drug resistance, decrease treatment duration, and adverse effects. By modulating the host immune response, HDT ameliorates immunopathological damage and improves overall outcomes by promoting autophagy, antimicrobial peptide production, and other mechanisms. It holds promise for addressing the challenges posed by multiple and extensively drug-resistant *Mycobacterium tuberculosis* strains, which are increasingly difficult to treat using conventional therapies. This article reviews various HDT candidates, including repurposed drugs, explores their underlying mechanisms such as autophagy promotion and inflammation reduction, while emphasizing their potential to improve TB treatment outcomes and outlining future research directions.

Keywords *Mycobacterium tuberculosis*, Tuberculosis, Infectious diseases, Host response, Host-directed therapy

Introduction

Tuberculosis (TB) is a chronic infectious disease caused by *Mycobacterium tuberculosis* (*M.tb*) and is the second leading cause of death from infectious diseases worldwide, following coronavirus disease 2019. TB remains a global public health issue, posing a serious threat to human health. The World Health Organization 2022 report estimates 10.6 million new cases of TB and 1.6 million associated deaths [1]. Currently, the main treatment regimen for TB involves a multidrug chemotherapy administered for at least six months. However, this regimen often involves prolonged courses of multiple antibiotics, which can lead to poor patient compliance, the

emergence of multidrug-resistant TB (MDR-TB), and adverse reactions [2]. Therefore, there is a pressing need to explore alternative therapeutic approaches.

Host immune response is critical to the pathogenesis of TB. As a novel therapeutic approach, host-directed therapy (HDT) has gained attention as an avenue for improving treatment outcomes of drug-resistant TB and is being increasingly explored as an adjunct TB treatment [3]. By focusing on the intricate interactions between *M.tb* and host immune cells, HDT exploits these metabolic relationships to bolster protective mechanisms against the pathogen. During TB infection, HDT modulates host cell functions, enhances protective immune responses, and improves the mycobacterial killing activities of host immune cells, working to eradicate or limit TB infection [4, 5]. Conversely, HDT also balances immune reactivity by reducing exacerbated inflammation and tissue damage associated with TB infection through precise regulation of the host immune system [6]. This approach eliminates *M.tb* through mechanisms that bypass the pathways commonly targeted by conventional anti-TB drugs, thereby reducing the risk of developing antibiotic resistance [5, 7]. This is particularly beneficial given the

[†]Na Tian and Hongqian Chu have contributed equally to this work.

*Correspondence:

Naihui Chu
dongchu1994@sina.com
Zhaogang Sun
sunzhaogang@bjxkyy.cn

¹ Department of Tuberculosis, Beijing Tuberculosis and Thoracic Tumor Research Institute, Beijing 101149, China

² Translational Medicine Center, Beijing Chest Hospital, Capital Medical University, Beijing 101149, China



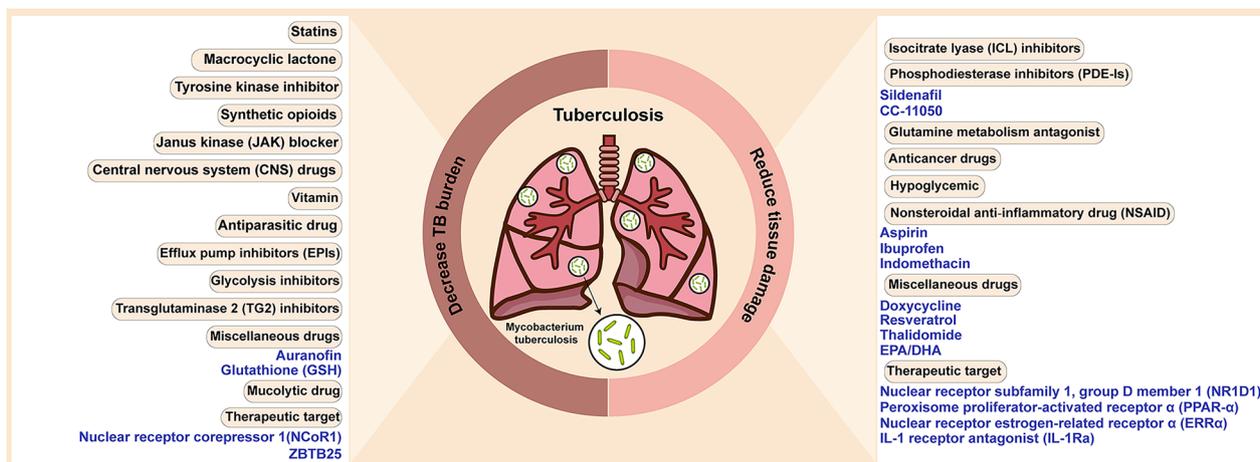


Fig. 1 Main current HDTs used in tuberculosis

rise of multidrug-resistant (MDR) and extensively drug-resistant (XDR) tuberculosis strains. This review focuses on a few drug candidates for HDT from two different perspectives: decreasing bacterial burden and mitigating pathological inflammatory responses (Fig. 1). Some of these drugs have been approved for the treatment of other diseases, such as simvastatin, everolimus, gefitinib and so on. In conducting our literature search, we aimed to comprehensively cover all significant HDT approaches for TB that have been reported over the years. While our search was not restricted to a specific time window, we focused on identifying key studies and advancements that have shaped the development of HDTs for TB. However, given the vast and evolving nature of the field, it is possible that some studies may not have been included. To ensure a thorough review, we utilized multiple databases, including PubMed, Scopus, and Web of Science, and employed broad search terms related to HDTs and TB. This approach allowed us to capture a wide range of studies, from early foundational work to recent clinical trials, providing a balanced overview of the field.

In addition, we have provided a partial list of these promising HDT agents and their targets of action (Table 1).

Promising candidates that decrease TB burden

Statins

Statins, specifically simvastatin and rosuvastatin, are widely used to lower circulating cholesterol levels and prevent coronary heart disease. These drugs exhibit anti-inflammatory and immunoregulatory properties while promoting cell autophagy and phagosome maturation by inhibiting the mammalian target of the rapamycin (mTOR) signaling pathway [8]. Autophagy, an

intracellular process that catabolizes intracellular components through lysosomal degradation, plays a crucial role in the host defense against intracellular pathogens like *M.tb* [57] (Fig. 2). Macrophages are key immune cells that inhibit *M.tb* growth via autophagy and facilitate the presentation of antigens to other immune cells.

Preclinical studies in *in vitro* models have shown that simvastatin exerted its anti-tubercular activity through cholesterol-driven autophagy, which is mediated by the adenosine monophosphate-activated protein kinase (AMPK)–mammalian target of the rapamycin complex 1 (mTORC1)–TFEB axis [58] (Fig. 3). In human peripheral blood mononuclear cells (PBMCs), simvastatin significantly reduced *M.tb* growth and promoted apoptosis and autophagy [59]. Furthermore, statins like simvastatin, fluvastatin, and pravastatin improved the efficacy of anti-TB drugs in cell models. In animal models, pravastatin demonstrated strong adjuvant action in a mouse model of human-like necrotic TB lung granulomas [60]. These preclinical findings suggest that statins have potential as adjunctive therapies in TB treatment. In contrast, clinical trials have yielded mixed results. A phase IIA study of atorvastatin showed a significant reduction in mycobacterial load in the sputum of patients with pulmonary tuberculosis (PTB) [61], while another trial using rosuvastatin did not affect sputum culture conversion [62]. These results suggested that not all statins are equally effective in clinical settings, and further research into optimal dosing and specific statin classes is warranted.

Macrocyclic lactone

Rapamycin, a macrocyclic lactone isolated from *Streptomyces hygroscopicus*, is known for its immunosuppressive and antimicrobial properties due to its

Table 1 Emerging HDTs against TB

Therapeutic class	Candidates	Target	References
Statins	Simvastatin	Autophagy	[8]
Macrocyclic lactone	Everolimus	Inhibiting the mTOR pathway; Autophagy	[9]
Tyrosine kinase inhibitor	Gefitinib	Tyrosine-kinase inhibitors (TKI); Autophagy	[10, 11]
Synthetic opioids	Loperamide	Autophagy; LL37 expression	[12, 13]
Janus kinase (JAK) blocker	Tofacitinib	JAK blocker	[14]
Central nervous system (CNS) drugs	Fluoxetine	Selective serotonin reuptake inhibitor (SSRI)	[15]
Vitamin	Vitamin D	LL37 expression; Autophagy; Macrophage polarization	[16–18]
Antiparasitic drug	Nitazoxanide (NTZ)	Autophagy	[19–21]
Efflux pump inhibitors (EPIs)	Verapamil	EPI	[22]
Glycolysis inhibitors	2-deoxyglucose (2-DG)	Glucose uptake inhibitor	[23]
Transglutaminase 2 (TG2) inhibitors	Cystamine and cysteamine	Reducing agents	[24]
Miscellaneous drugs	Auranofin	Thioredoxin reductase (TrxR) inhibition	[25]
	Glutathione (GSH)	Th1 response regulation	[26]
Mucolytic drug	Ambroxol	Autophagy	[27]
Therapeutic target	Nuclear receptor corepressor 1 (NCOR1)	Autophagy	[28]
	Host transcriptional repressor protein zinc finger and BTB domain 25(ZBTB25)	Autophagy	[29]
Isocitrate lyase (ICL) inhibitors	Itaconate	ICL inhibitor	[30–32]
Phosphodiesterase inhibitors (PDE-Is)	Sildenafil	PDE-I	[33, 34]
	CC-11050	PDE-I	[35]
Glutamine metabolism antagonist	JHU083	Glutamine metabolism antagonist	[36]
Anticancer drugs	Bevacizumab (Avastin)	Anti-vascular endothelial growth factor (VEGF) antibody	[37]
Hypoglycemic	Metformin	Adenosine monophosphate-activated protein kinase (AMPK) activation; Autophagy	[38–40]
Nonsteroidal anti-inflammatory drug (NSAID)	Aspirin	Cyclooxygenase enzymes inhibitor	[41]
	Ibuprofen	Cyclooxygenase enzymes inhibitor	[42, 43]
	Indomethacin	Cyclooxygenase enzymes inhibitor	[44]
Miscellaneous drugs	Doxycycline	Matrix metalloproteinase (MMP) inhibitor	[45, 46]
	Resveratrol	Sirtuin (Sirt1) activation	[47, 48]
	Thalidomide	TNF- α inhibition	[49]
	EPA/DHA	Inflammation regulation	[50–52]
Therapeutic target	Nuclear receptor subfamily 1, group D member 1 (NR1D1)	Phagosome lysosome maturation	[53]
	Peroxisome proliferator-activated receptor α (PPAR- α)	Autophagy	[54]
	Nuclear receptor estrogen-related receptor α (ERR α)	Autophagy	[55]
	IL-1 receptor antagonist (IL-1Ra)	Blocking signaling	[56]

ability to inhibit the mTOR pathway, thereby promoting autophagy [63]. Specifically, rapamycin binds to FKBP12, forming a complex that inhibits mTORC1 activity. This inhibition relieves the suppression of ULK1, a key initiator of autophagy, leading to the

formation of autophagosomes [64]. Preclinical studies in *M.tb*-infected mice have shown that rapamycin reduced pulmonary inflammation and caseating granulomas [65]. However, its application in clinical settings is restricted by high variability in absorption and

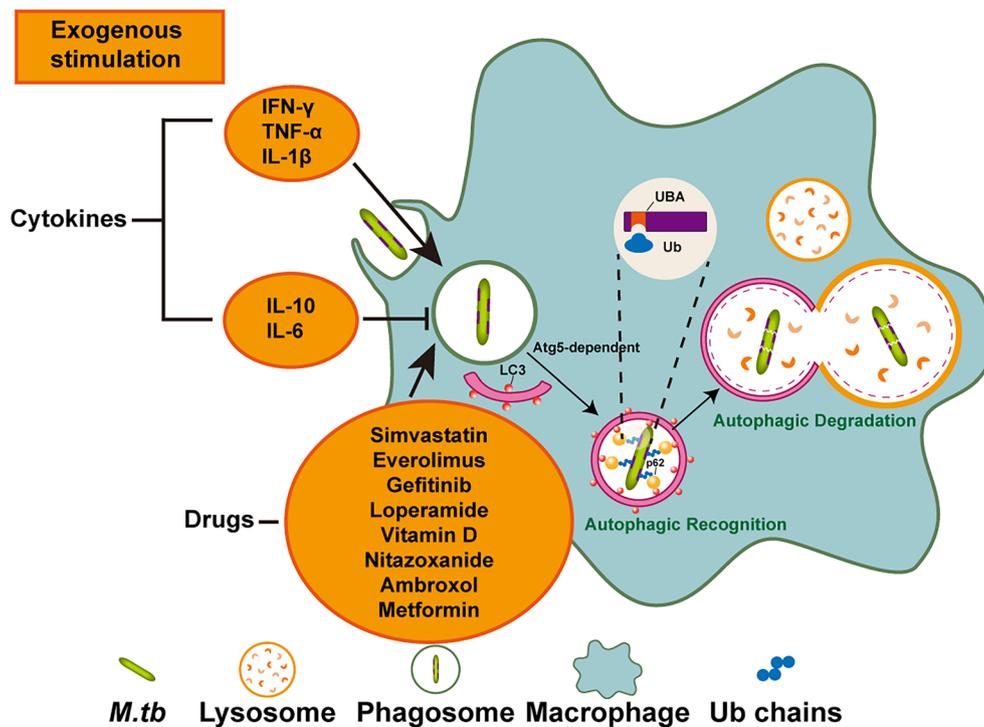


Fig. 2 Schematic representation of the correlation between autophagy induction and the anti-tuberculosis activity of indicated cytokines and pharmacological agents

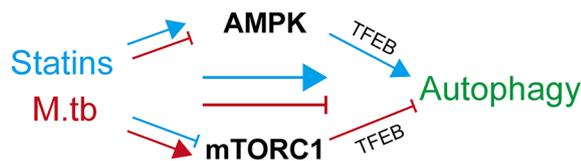


Fig. 3 Statins and *M.tb* infection exhibit opposing effects on AMPK, mTORC1, and autophagy. Statins suppress mTORC1 activity while activating AMPK, both of which cause enhanced nuclear translocation of TFEB, resulting in the expression of autophagy-related genes. *M.tb* produces opposite effects by activating mTORC1 and blocking AMPK, which prevents nuclear translocation of TFEB and inhibits autophagy

It modulates cytokine profiles and oxidative stress, further supporting its role in HDT [66]. Moving to clinical outcomes, a phase II trial investigating everolimus as an adjunctive therapy for TB showed that it was well tolerated and might enhance recovery of lung function, particularly in terms of FEV₁ [67]. While everolimus has shown promise in modulating the immune response and improving TB treatment outcomes, there are significant concerns about long-term use due to its immunosuppressive effects. Further research should assess the balance between its potential benefits and risks, particularly in patients with concurrent infections or immunocompromised states.

adverse reactions. Rapamycin's efficacy is further limited due to its metabolism via cytochrome P450 3A4 (Cyp3A4), which is induced by rifampin, complicating its use alongside standard TB treatments.

Everolimus, a derivative of rapamycin with better pharmacokinetics, has shown promising results in pre-clinical studies. At the cellular level, everolimus has been demonstrated to enhance autophagy by inhibiting PI3K/Akt/mTOR pathway. In in vitro granuloma models, everolimus not only controlled *M.tb* growth but also demonstrated an additive effect when combined with front-line anti-TB drugs like isoniazid and pyrazinamide.

Tyrosine kinase inhibitor (TKI)

Imatinib, a TKI known for its antitumor activity, has shown promise in interfering with the entry and survival of *M.tb* in macrophages. Preclinical studies in *M.tb*-infected mice have demonstrated that imatinib decreased granulomatous lesions and bacterial load. It also demonstrated efficacy against antibiotic-resistant strains by promoting autophagy, potentially contributing to *M.tb* clearance and reducing the risk of drug-resistant diseases [68]. Another TKI, gefitinib (ZD1839, Iressa), which targets the epidermal growth factor receptor (EGFR), has

been shown to restrict *M.tb* replication in both macrophages as well as in the lungs of *M.tb*-infected mice by promoting the host autophagy pathway through the inhibition of the p38 MAPK signaling pathway activated by EGFR [10]. Additionally, gefitinib has also been shown to control *M.tb* growth in macrophages by enhancing lysosomal biogenesis and function through the inhibition of the signal transducer and activator of transcription 3 (STAT3) signaling pathway [11]. However, further clinical trials are necessary to evaluate the safety and efficacy of TKI as adjunctive therapies in TB treatment, with a focus on understanding how TKI might be safely integrated into clinical practice.

Synthetic opioids

Loperamide, a synthetic opioid routinely prescribed to treat diarrhea, demonstrates potential antimicrobial effects against *M.tb*. Preclinical studies in murine and human alveolar macrophages (AMs) have demonstrated that loperamide induced autophagy, enhancing its antimicrobial activity towards *M.tb* [12]. In addition, loperamide was found to reduce TNF- α levels and induce the production of antimicrobial peptides, such as bactericidal/permeability increasing protein and LL37, enhancing host's ability to fight against *M.tb* infections while protecting tissues from excessive inflammatory damage [13]. However, as an opioid, loperamide has potential side effects, including the risk of opioid dependence and gastrointestinal disturbances. Further clinical trials are necessary to determine the appropriate dosage, efficacy, and safety of loperamide in patients with PTB. Additionally, studies should explore whether loperamide's immunomodulating effects can be harnessed without inducing opioid-related side effects.

Janus kinase (JAK) blocker

Tofacitinib, a JAK blocker, accelerated bacterial elimination in a preclinical study when administered at high doses alongside standard TB chemotherapy in chronic *M.tb*-infected BALB/c mice. However, similar results were not observed in *M.tb*-infected C3HeB/FeJ mice with *Ipr1* gene mutations. This difference may be attributed to the necrotic and hypoxic granulomatous lesions in C3HeB/FeJ mice, which could hinder the action of tofacitinib. These findings highlighted the importance of host factors such as granuloma structure and immune environment in determining the efficacy of JAK blockers [14]. However, the potential immunosuppressive effects of JAK inhibitors also pose challenges, particularly in immunocompromised individuals or those with concurrent infections. Further studies are warranted to investigate its effectiveness as an adjuvant treatment during the

early stages of infection (approximately 3 weeks), prior to the onset of pathological necrosis.

Central nervous system (CNS) drugs

Dopamine and serotonin receptors mediate the immune responses to TB. Preclinical studies have shown that agonists or antagonists of these receptors activated autophagy, thereby enhancing the host's ability to eliminate *M.tb*. For example, the combination of fluoxetine (a 5-HT_{2C} receptor antagonist) and bromocriptine (an agonist of type 2 dopamine receptors) was found to increase the release of proinflammatory cytokine (IL-6 and TNF- α) and stimulate autophagy in *M.tb*-infected macrophages [10, 15]. A synthetic fluoxetine analog (AM3e) also exhibited anti-TB efficacy against H37Rv strain [69]. While CNS drugs like sertraline (SRT) have shown promise in animal models, where their combination with front-line anti-TB drugs, improved early bactericidal activity, resolution of pulmonary pathology, and enhanced survival in mice infected with *M.tb* [70], clinical studies have raised concerns. For instance, a recent clinical trial on SRT as an adjunct regimen for asymptomatic cryptococcal antigenemia reported serious adverse events (psychosis, aggressive behavioral changes, and serotonin syndrome) [71]. Although CNS drugs may enhance TB treatment through immune modulation, their use as adjunctive therapies must be approached cautiously due to the risk of severe side effects. Further studies are necessary to determine the optimal dosage and treatment duration to ensure patient safety.

Vitamins

Vitamin D, a fat-soluble secosteroid hormone, plays a key role in combating *M.tb* by impairing bacterial growth and upregulating innate host responses [72]. Its active form, 1,25(OH)₂D, regulates mucosal immunity, host defense, and inflammation by binding to the vitamin D receptor (VDR) in macrophages [73, 74], triggering the expression of the antimicrobial peptide cathelicidin (CAMP) (LL-37) and promoting autophagosome-lysosome fusion and maturation [16, 75]. Preclinical evidence shows that 1,25(OH)₂D induces a shift from inflammatory (M1) to reparative (M2) macrophages, promoting the secretion of hydrogen peroxide, a critical factor for *M.tb* clearance [17, 18]. Clinical trials showed that cholecalciferol (vitamin D) supplementation, especially in patients with TB having 1,25(OH)₂D deficiency, may augment standard anti-TB therapy (ATT) by accelerating sputum culture conversion and improving lesion absorption [7, 76]. However, other studies have reported mixed results, with some trials showing no significant impact on TB relapse or culture conversion [77, 78]. These findings underscore

the need for larger multicenter trials to fully assess the clinical utility of vitamin D as an adjunct therapy for TB.

Nitazoxanide (NTZ)

Nitazoxanide, an antiparasitic drug with notable antiviral and anti-inflammatory properties [79]. In preclinical studies, NTZ suppresses intracellular *M.tb* proliferation by inhibiting the enzymatic activity of human quinone oxidoreductase (NQO1), which in turn inhibits the mTORC1 pathway and induces autophagy [19]. In vivo studies demonstrated that NTZ, when combined with INH and rifabutin (RFB), successfully cleared all bacteria from the lungs and spleens of *M.tb*-infected mice and significantly restored tissue architecture [21]. However, a phase II clinical trial revealed that NTZ did not exhibit bactericidal activity against *M.tb* in drug-susceptible patients with PTB. This lack of efficacy may be attributed to the low plasma and sputum concentrations of NTZ and the high plasma protein binding of nitazoxanide, the active metabolite of NTZ [80], highlighting the need for further pharmacokinetic optimization and clinical research to better understand its potential role in TB treatment.

Efflux pump inhibitors (EPIs)

Verapamil, a calcium channel blocker used to treat hypertension, angina, and cardiac arrhythmia [81], has been identified as an EPI with potential application in TB therapy. Firstly, verapamil inhibits drug efflux pumps in *M.tb*, such as Rv1258c and Rv2686c, thereby increasing the intracellular concentration of anti-TB drugs like rifampicin and isoniazid. This not only enhances drug efficacy but also reduces the likelihood of drug resistance [82]. Secondly, Verapamil modulates host immune responses by promoting macrophage activation and increasing the production of reactive oxygen species (ROS) and nitric oxide (NO), which contribute to the killing of intracellular mycobacteria [83]. Additionally, Verapamil has been shown to reduce bacterial persistence, a major challenge in TB treatment, by enhancing drug accumulation and immune-mediated clearance [84]. In vitro studies demonstrated that verapamil, when combined with rifapentine, enhanced antimicrobial activity against *M.tb* in macrophages [85]. Animal models showed that verapamil accelerated bactericidal activity and achieved durable sterilization in infected mice [86]. However, verapamil's dual effects on drug metabolism, particularly its inhibition of CYP3A activity, raise concerns about potential interactions with essential TB drugs like rifampin, which is known to induce CYP3A enzymes [87]. Therefore, further clinical investigations are needed to determine the impact of verapamil on

cardiac conduction, drug metabolism, and its potential as an adjunctive therapy for TB before regulatory agency clearance.

Glycolysis inhibitors

The Warburg effect, initially observed in cancer cells, refers to a metabolic state in which cells favor aerobic glycolysis over oxidative phosphorylation for ATP and macromolecule production. This phenomenon has also been observed in the immune cells during *M. marinum* infection. Preclinical studies demonstrated that pretreatment with 2-deoxyglucose (2-DG) induced autophagy and restricted the growth of *M. marinum* in zebrafish larvae, potentially enhancing host defenses against mycobacterial infections. However, post-infection treatment with 2-DG failed to inhibit bacterial replication and even promoted *M. marinum* growth in TNF- $\alpha^{-/-}$ zebrafish [23]. This was consistent with another study showing that 2-DG treatment of *M.tb*-infected macrophages enhanced *M.tb* growth [88]. While clinical studies have established that 2-DG can be safely administered to humans [89], its potential application in TB treatment is limited, particularly due to its effectiveness only when administered before infection. The potential of glycolysis inhibitors in post-infection TB therapy warrants further investigation, but current evidence suggests significant challenges in their use as therapeutic agents.

Transglutaminase 2 (TG2) inhibitors

Cystamine and cysteamine, as TG2 inhibitors, function as reducing agents that elevate the levels of glutathione and L-cysteine [90], thereby impacting cell metabolism. Preclinical studies demonstrated that these agents restricted *M.tb* growth in macrophages and suppressed host cell autophagy but did not exhibit direct bactericidal activity against *M.tb* cultures. In vitro models indicated that combining TG2 inhibitors with amikacin enhanced their antimicrobial activity in *M.tb*-infected macrophages and the granuloma-like structure ex vivo model [24]. These results showed that their combination with existing antibiotics like amikacin may offer potential, but clinical trials are essential to validate their efficacy and safety in TB patients.

Miscellaneous drugs

Auranofin

Auranofin, an orally administered anti-rheumatic drug, exhibits broad-spectrum antibacterial and antiviral activity in vitro. Preclinical studies indicated that it targeted the bacterial flavoenzyme TrxR, essential for *M.tb* survival by protecting against oxidative and nitrosative stress. By inhibiting TrxR, auranofin compromises *M.tb* defense mechanisms, especially in the

oxidative environment of macrophage phagosomes [91]. Despite showing efficacy in ex vivo whole blood cultures, aurano-fin did not affect sputum culture conversion in early clinical trials [25]. Its potential against acute and latent *M.tb* infections is still under investigation, and small-scale clinical trials may be necessary to assess its anti-TB efficacy either as a monotherapy or in combination with other anti-TB drugs.

Glutathione (GSH)

GSH, a naturally occurring antioxidant, plays a vital role in protecting cells from oxidative stress-induced damage, regulating DNA expression, detoxifying reactive metabolites, providing cysteine reservoirs, modulating apoptosis and antigen-presenting cell functions [92]. In vitro studies showed that H37Rv strain is sensitive to GSH, highlighting its potential for intracellular *M.tb* control [93]. Liposomal glutathione (L-GSH) supplementation was found to decrease the intracellular mycobacterial burden within in vitro granulomas derived from the PBMCs of patients with type 2 diabetes mellitus (T2DM) [26]. However, the small sample size and in vitro granuloma model used thus far do not fully replicate the complex granulomatous environments observed in PTB patients. Further small-scale clinical trials are needed to validate GSH's therapeutic effects in patients with TB.

Ambroxol (Amb)

Ambroxol, an active metabolite of bromhexine, is approved for the treatment of airway diseases in several countries. Preclinical studies have demonstrated that ambroxol enhanced the antimycobacterial activity of rifampicin against intracellular BCG [27], induced autophagy through the activation of TFEB both in vitro and in vivo models, promoted mycobacterial killing in macrophages. Moreover, it has been shown to potentiate rifampin's activity in a murine tuberculosis model [94]. However, the optimal dosage of Amb when co-administered with rifampin or vancomycin remains under investigation. Amb is metabolized by Cyp3A4, which is strongly induced by rifampicin in humans. High-dose Amb (1000 mg/day) has been used clinically to counteract rifampicin-induced Cyp3A4 activation and achieve sufficient Amb levels. However, this increased dose also resulted in higher levels of Cyp3A4-derived Amb metabolites, whose clinical implications remain unclear. Further clinical trials are needed to explore the interaction of Amb with other TB drugs and long-term impact on patients.

Therapeutic target

Nuclear receptor corepressor 1 (NCoR1)

NCoR1 is a scaffolding protein that forms the foundation of a large corepressor complex responsible for suppressing the expression of genes involved in various biological processes [95]. The NCoR1 corepressor is involved in both autophagy and lysosome biosynthesis by fine-tuning ATP homeostasis via the AMPK–mTOR–TFEB signaling axis to control *M.tb* infection in myeloid cells. Preclinical studies have demonstrated that NCoR1 depletion, followed by treatment with rapamycin, anti-mycin-A, or metformin restored TFEB activity and LC3 levels, leading to improved clearance of *M.tb* [28]. Thus, NCoR1 is a promising therapeutic target for HDT in TB. However, it remains to be determined whether its beneficial effects are primarily mediated via the inhibition of direct PPAR γ target genes, such as CD36, or through the PPAR γ -mediated repression of proinflammatory transcription factors, such as NF- κ B.

ZBTB25

ZBTB25, a host transcriptional repressor protein associated with the histone deacetylase 1 (silencing complex), has been identified as another potential target for HDT in TB. Preclinical evidence has shown that knock-down of ZBTB25 enhanced the release of IL-12p40 from infected macrophages. Treatment of macrophages with ZBTB inhibitors further induced autophagy and eliminated intracellular *M.tb* by enhancing JAK2 and STAT4 phosphorylation [29]. Additional research is necessary to determine the efficacy and safety of ZBTB inhibitors in clinical trials, as well as their potential in combination therapies with existing anti-TB drugs.

Promising candidates that reduce tissue damage

Tuberculoid granulomas (Fig. 4), the hallmark lesions of chronic TB, are well-organized, compact structure composed of differentiated macrophages, lymphocytes, and other immune cells [96, 97]. As granulomas mature, they develop fibrous walls and experience a significant decrease in the number of vessels penetrating the structure, which hampers the penetration of anti-TB drugs into the necrotic and hypoxic areas of the granuloma, where *M.tb* persists [98, 99]. The poor permeability of anti-TB drugs within granulomas, coupled with ineffective regulation of extracellular signaling, allows *M.tb* to adapt and develop drug resistance. In such environments, *M.tb* switches to fatty acid metabolism to enter a persistent state. HDT strategies can improve therapeutic outcomes by enhancing immune homeostasis during the formation and resolution of granulomas. By improving vascular perfusion within granulomas, HDT can increase

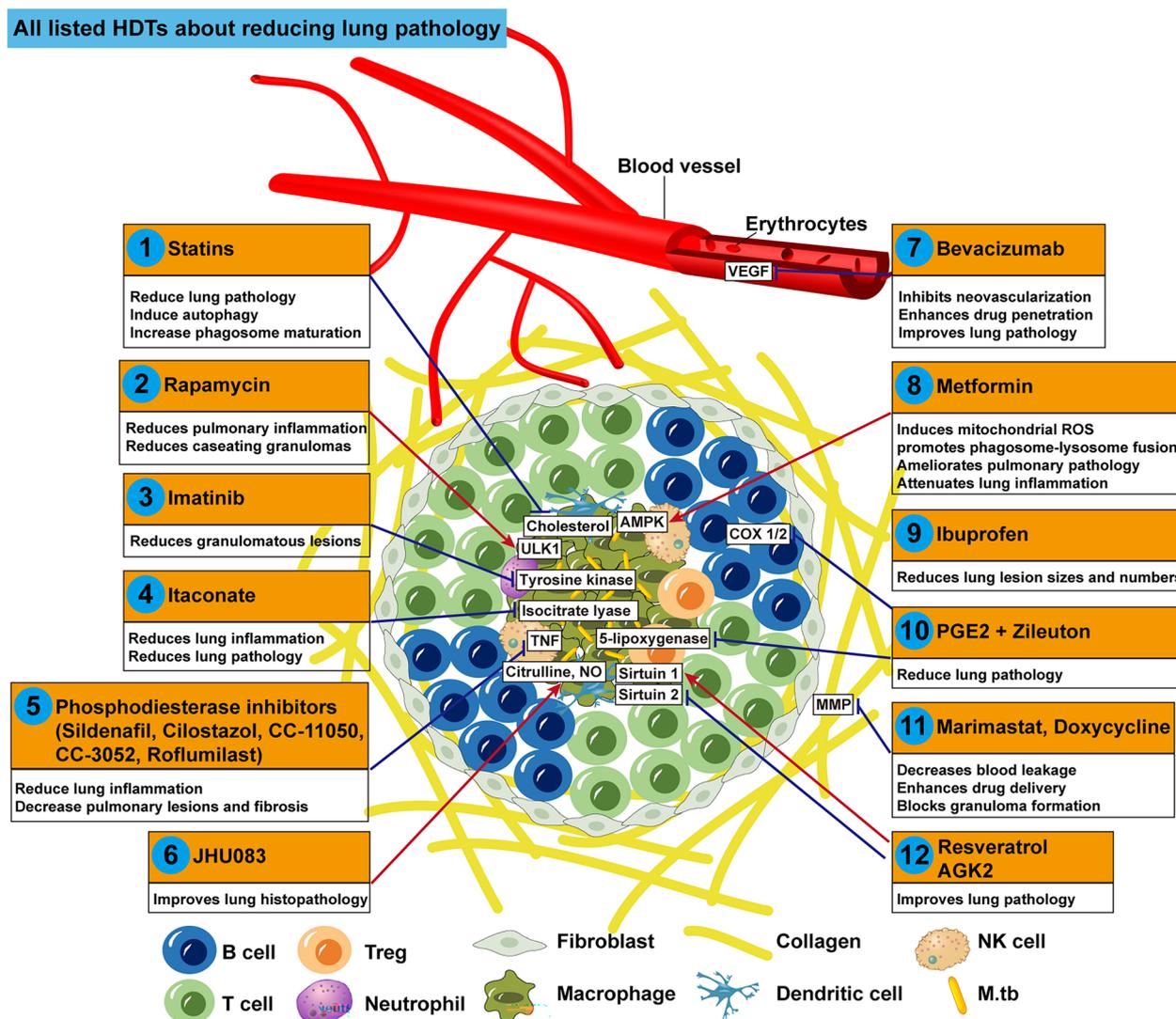


Fig. 4 The structure and main composition of granuloma and related drugs

the local concentration of antimicrobials, promote tissue repair, and enhance the clearance of latent mycobacteria by immune cells.

Isocitrate lyase (ICL) inhibitors

ICL plays a crucial role in mycobacterial glyoxylate and methyl isocitrate cycles, which are essential for *M.tb* survival, virulence [100], and antibiotic tolerance [101]. Preclinical studies demonstrated that itaconate, a structural analog of succinate, inhibits ICL through covalent interaction with the catalytic cysteine residues (Cys191 of ICL1 and Cys215 of ICL2) of *M.tb* [30]. Dimethyl itaconate activates innate immune defenses, maintains inflammatory homeostasis, activates STAT3 and autophagy, and exhibits strong antimycobacterial

properties in both macrophages and in vivo models [102]. Additionally, itaconic acid promotes the pentose phosphate pathway, leading to ROS production, enhancing its anti-inflammatory and antibacterial properties [103]. In addition, Irg1-deficient phagocytes, which cannot produce itaconate, have shown increased susceptibility to *M.tb* infection, with infected mice exhibiting higher bacterial loads, reduced survival, exacerbated inflammation, extensive necrotizing granuloma formation and elevated neutrophil infiltration in the lungs [104, 105]. In summary, the development of pharmacological agents that promote itaconate production could potentially mitigate pathological inflammatory responses and prevent severe lung injury

associated with TB progression. But further research is needed to confirm these findings in clinical trials.

Phosphodiesterase inhibitors (PDE-Is)

Phosphodiesterases (PDEs) regulate the concentration of cyclic adenosine monophosphate (cAMP), a key second messenger in both eukaryotic and prokaryotic systems [106]. *M.tb* manipulates cAMP signaling pathways within infected macrophages, thereby influencing its survival [107]. Preclinical studies have showed that PDE inhibitors, such as sildenafil and cilostazol, can decrease tissue pathology, accelerate bacterial clearance, and reduce the time to lung sterilization in *M.tb*-infected mice when used in combination with standard TB therapy [33]. PDE4 inhibitors, including CC-11050, CC-3052, and roflumilast, when combined with INH, have been shown to reduce the lung bacillary burden, lung pathology, and fibrosis, as well as the size and number of lung granulomas in *M.tb*-infected rabbits and in both acute and chronic TB mouse models [35, 108, 109]. A phase 2 clinical trial demonstrated that CC-11050 and everolimus were safe and well tolerated as TB adjuvant treatments and enhanced recovery of lung function (FEV₁), which correlates with reduced all-cause mortality [67]. However, contrasting data from other studies showed that the addition of PDE4i rolipram to standard TB treatment accelerated mortality, increased the bacterial burden, and did not reduce the time to bacterial clearance in the lungs of a mouse model of TB [34], raising concerns about the general applicability of PDE-Is. Therefore, understanding the differential impacts of various PDE-Is and their long-term outcomes is essential before widespread clinical use.

Glutamine metabolism antagonist

Glutamine metabolism is crucial for T-cell cytokine production and M1-like polarization of macrophages in the proinflammatory response against *M.tb* infection [110]. Glutamine also serves as a key carbon and nitrogen source for *M.tb*-infected macrophages [111], supporting the pathogen's metabolic demands. The glutamine metabolism antagonist, JHU083, has been shown to inhibit *M.tb* replication both in vitro and in vivo, improving survival, reducing lung bacterial burden, and improving lung histopathology. These effects are achieved by reducing immunosuppressive myeloid cells, increasing effector T cells, and enhancing the production of citrulline and NO. However, in *M.tb*-infected immunocompromised mice, JHU083 showed reduced therapeutic efficacy [36]. This suggests that its benefits may be limited in patients with weakened immune systems, such as those co-infected with HIV. Therefore, further clinical trials are needed to assess its potential as an immunotherapeutic

agent against TB, particularly in diverse patient populations with varying immune statuses.

Anticancer drugs

Polena et al. demonstrated that blocking vascular endothelial growth factor (VEGF) signaling with antibodies against VEGF or VEGFR-2 reduced *M.tb* transmission to the lung, spleen, and liver in *M.tb*-infected mice [112]. In line with this, bevacizumab (Avastin), a humanized monoclonal antibody against VEGF commonly used in cancer treatment, has demonstrated potential in the context of TB. In *M. marinum*-infected zebrafish, targeting VEGFR signaling improved the therapeutic effect of rifampicin [113]. Additionally, a preclinical study using a rabbit TB model showed that bevacizumab inhibited neovascularization, enhanced drug penetration into granulomas, improved lung pathology, and increased oxygenation, suggesting its potential to enhance the effectiveness of the current TB regimens [37]. However, larger multicenter randomized controlled trials are required to formulate guidelines for the clinical use of bevacizumab in treating tubercular granulomas. VEGF-A, which binds to VEGFR1, functions as an effective chemokine for macrophages. According to Harding et al., suppressing VEGF-A reduced granulomatous inflammation by interfering the recruitment of monocytes to infected tissues [114]. This suppression could potentially improve the survival of mice infected with virulent *M.tb*, while maintaining host defense. Thus, VEGF-A presents a potential target for therapeutic interventions in TB therapy.

Metformin

Metformin, a first-line drug for patients with T2DM [115], is known to activate AMPK [38], a key metabolic regulator. AMPK promotes autophagy by inhibiting mTOR, a potent negative regulator of autophagy, or by activating unc-51-like kinase 1, which is essential for autophagy initiation [39, 40]. Furthermore, the AMPK/PPAR γ coactivator-1 α (PGC1 α) pathway upregulates autophagy-related genes, thereby promoting autophagy and phagosome fusion in macrophages during *M.tb* infection [116]. In vitro research found that metformin activated a novel galectin-directed ubiquitin signal transduction system in response to lysosomal membrane damage, induces autophagy, and facilitates the elimination of *M.tb* from macrophages [117]. Additionally, metformin inhibits *M.tb* growth, induces mitochondrial ROS production, promotes phagosome-lysosome fusion in vitro, ameliorates pulmonary pathology, attenuates chronic inflammation, augments immunity, and improves the therapeutic effect of the standard anti-TB regimen in *M.tb*-infected mice. In clinical studies, metformin therapy was associated with improved clinical outcomes,

including reduced TB severity, and a lower incidence of latent TB infection (LTBI) [118]. Retrospective studies also suggested that metformin decreased the risk of TB incidence [119, 120], cavitary TB [118], and mortality associated with diabetes mellitus during TB treatment [121]. Additionally, metformin has been shown to improve sputum culture conversion and lower the relapse rate of patients with TB-DM [122]. However, in one study, the use of a low dosage of metformin in combination with conventional ATT did not accelerate sputum culture conversion, which could be attributed to insufficient dosing [123]. Despite this, the role of metformin in reducing inflammation and improving overall outcomes makes it a promising adjunctive therapy in HDT for TB. Further prospective clinical trials are needed to determine whether the initial use of metformin can effectively prevent TB infection after exposure or simply mitigate the progression from LTBI to active disease. These studies should also aim to establish the optimal dose when metformin is used alongside current TB therapies, as well as its potential role in shortening treatment duration and enhancing prophylactic control of TB.

Nonsteroidal anti-inflammatory drug (NSAID)

Aspirin

Aspirin, a commonly used NSAID, primarily works by inhibiting cyclooxygenase (COX) enzymes, particularly COX-1 and COX-2, which in turn blocks thromboxane A₂ synthesis [124]. A study conducted on *M.tb*-infected mice showed that the co-administration of aspirin and pyrazinamide enhanced the efficacy of pyrazinamide during the early stage of TB treatment [42]. Additionally, a phase 2, randomized placebo-controlled trial showed that aspirin reduced stroke and death in HIV-uninfected adults with TB meningitis (Table 2) [41]. However, given the widespread use of aspirin, which may overlap with anti-TB treatment, it is necessary to determine their potential interaction in vivo. Initial assessment should be conducted using animal models, followed by larger studies involving children and adults. In addition, long-term TB treatment studies in mice, evaluating the effects of aspirin in combination with pyrazinamide and other TB drugs are needed to determine the potential role of aspirin in reducing TB treatment and preventing relapse.

Ibuprofen

Ibuprofen, another commonly used NSAID, inhibits COX-1 and COX-2 [125], both of which are involved in the synthesis of prostaglandin E₂ (PGE₂) in macrophages [126]. COX inhibition has been shown to enhance nitric oxide synthase (iNOS), TNF- α and IFN- γ expression and reduce mycobacterial loads by inhibiting the expressions of the PGE₂ [127]. However, PGE₂ is known to play a

dual role, as it can both suppress the immune response against *M.tb* and protect the host from excessive inflammation, suggesting that NSAIDs should be exercised with caution for TB treatment [128]. In some studies, the 5-lipoxygenase inhibitor zileuton, when co-administered with PGE₂, reduced pulmonary bacterial burden and pathology and prevented acute death in *M.tb*-infected mice deficient in both IL1R1 and IFNAR1 [129]. Early animal studies also reported that ibuprofen enhanced the mycobactericidal effect of pyrazinamide in *M.tb*-infected mice [42], reducing the size and number of lung lesions, decreasing bacterial load, and prolonging survival of C3HeB/FeJ mice, which are particularly vulnerable to TB [43]. However, the combination of ibuprofen with standard chemotherapy has not yet been evaluated. Additionally, some studies have raised concerns, as ibuprofen treatment can increase the bacterial load and decrease survival in CB6F1 mice following high-dose aerosol infection [130]. Therefore, further animal experiments and clinical trials are needed to evaluate the effects of ibuprofen on both active and latent TB.

Indomethacin

Indomethacin, an NSAID that participates in both innate and adaptive immunity [131], has been shown to reduce the proportion of *M.tb* antigen-triggered Tregs, reactions of *M.tb* specific cytokines, and proliferation of T cells in active TB. This suggests their potential use in regulating immune responses against TB infection [44]. Nevertheless, future clinical trials should be conducted to explore the in vivo effects of indomethacin in patients during various stages of *M.tb* infection.

Miscellaneous drugs

Matrix metalloproteinase (MMP) inhibitor

MMP is a proteolytic enzyme that degrades the extracellular matrix, including elastin and collagen [136]. Elevated MMP levels are associated with lung inflammation and granuloma necrosis in TB. Marimastat (BB-2516), a small molecule MMP inhibitor, has shown promise by enhancing the in vivo efficacy of INH and RFP, increasing the proportion of healthy blood vessels at the infection site, and enhancing drug delivery and retention [137]. In a human lung tissue model of TB granulomas, marimastat blocked both granuloma formation and mycobacterial growth [45]. Moreover, anti-MMP-9 antibody treatment in C3HeB/FeJ mice led to the development of hypoxic TB granulomas and cavitary lesions, resulting in a lower relapse rate, although this was not significantly different [138]. Doxycycline, another MMP inhibitor, effectively prevented collagen destruction induced by TB infection [46]. Promising results from a recent phase II clinical trial showed that adjunctive doxycycline with standard

Table 2 Clinical studies of host-directed therapeutics for tuberculosis

Study name	Phase	Results	Clinical trials.Gov Identifier*	References
A Pilot Study of Adjunctive Aspirin for the Treatment of HIV Negative Adults With Tuberculous Meningitis	II	The addition of aspirin to dexamethasone may improve outcomes from tuberculous meningitis (TBM)	NCT02237365	[41]
Treating Tuberculosis With the Lipid Lowering Drug Atorvastatin in Nigeria (ATORvastatin in Pulmonary Tuberculosis)	II	Atorvastatin is a safe additive to standard anti-TB regimen and leads to a significant reduction in mycobacterial load in the sputum	NCT04721795	[61]
Rosuvastatin Evaluation as a Tuberculosis Treatment Adjunct	II	Adjunctive rosuvastatin at 10 mg once per day was safe but did not produce substantive benefits on culture conversion in the overall study population	NCT04504851	[62]
TB Host-Directed Therapy	II	CC-11050 and everolimus were safe and reasonably well tolerated as adjunctive therapies for tuberculosis, and analysis of preliminary efficacy suggests they might also enhance the recovery of FEV ₁	NCT02968927	[67]
A 14 Day Early Bactericidal Activity Study of Nitazoxanide for the Treatment of Tuberculosis	II	At the doses used, NTZ did not show bactericidal activity against M. tuberculosis	NCT02684240	[80]
Vitamin D Supplementation Effect In Children With Pulmonary Tuberculosis Treatment	Not applicable	Vitamin D is beneficial in improving fever and cough resolution, and improving nutritional status in children with pulmonary TB and vitamin D insufficiency	NCT05073965	[132]
Doxycycline in Human Pulmonary Tuberculosis	II	Adjunctive doxycycline with standard anti-TB treatment suppressed pathological MMP in PTB patients	NCT02774993	[133]
Clinical Trial of Phenylbutyrate and Vitamin D in Tuberculosis (TB)	II	Adjunct therapy with PBA + vitD3 or vitD3 or PBA to standard short-course therapy demonstrated beneficial effects towards clinical recovery and holds potential for host-directed-therapy in the treatment of TB	NCT01580007	[134]
Immune Reconstitution in Tuberculosis Disease	II	Daily supplementation with vitD3 + PBA may ameliorate clinical TB symptoms and disease-specific complications, while the intervention had no effect on bacterial clearance in sputum	NCT01698476	[135]

* Further details for trial with NCT numbers can be accessed at <http://clinicaltrials.gov>

anti-TB treatment may hold potential [132]. However, cipelestat, a selective MMP inhibitor, paradoxically increased cavitation, immunopathology, and mortality in a murine model of cavitary TB [139]. Further phase 3 clinical trials with larger sample sizes are needed to thoroughly assess the immunopathological effects of MMP inhibitors like doxycycline on TB.

Resveratrol

Sirtuin 1 (Sirt1), an NAD-dependent deacetylase, inhibits apoptosis in mammalian cells. *M.tb* infection downregulates Sirt1 expression, which is crucial for negatively modulating inflammatory responses by inhibiting the activation of TAK1, MAPK and NF- κ B pathways and reducing IL-6 and TNF- α levels. Resveratrol treatment reverses this effect [47] and demonstrates the ability to decrease bacterial burden and improve lung pathology in *Mabc*-infected mice and zebrafish by activating Sirt3 [140]. Furthermore, resveratrol inhibited both *M.tb*-induced early and later apoptosis in macrophages, thereby markedly inhibiting intracellular *M.tb* growth [48]. Moreover, Sirt2 enhances macrophage activation in response to *M.tb* infection. Inhibition of Sirt2 by AGK2 restricted both drug-sensitive and drug-resistant *M.tb* strains, enhanced the efficacy of isoniazid, reduced the bacillary burden and improved disease pathology in *M.tb*-infected mice [141]. Sirt7 restricts intracellular *M.tb* growth by enhancing NO release from macrophages and NO-dependent apoptosis [142]. However, further studies on different experimental models, such as non-human primates, and with a range of other chemical inhibitors of sirtuins are required to fully explore the potential of sirtuin inhibitors as HDT for TB.

Thalidomide

Thalidomide, originally used as a sedative, hypnotic, and antiemetic, could promote host immunity against *M.tb* infection by increasing cytokines production and T lymphocyte proliferation [143]. When combined with antibiotics, thalidomide lowered TNF- α levels, reduced leukocytosis, and alleviated brain pathology, resulting in markedly improved survival in a rabbit model of acute *M.tb* CNS infection [144]. Clinical studies have shown that the addition of thalidomide to the anti-TB treatment regimen resulted in substantial clinical and neuroradiological improvement, with a favorable safety profile and good tolerability in patients with tuberculous meningitis (TBM) [49]. However, owing to concerns about its teratogenic and potentially mutagenic effects, its use in TB treatment remains experimental and requires further investigation.

Omega-3 long-chain polyunsaturated fatty acids (n-3 LCPUFA)

N-3 LCPUFA, found in oily fish and supplements such as eicosapentaenoic acid and docosahexaenoic acid, are essential for numerous physiological and biochemical processes, especially in inflammation modulation. Studies have shown that adding n-3 LCPUFA to TB treatment regimens reduced *M.tb* burden [50], decreased systemic and lung inflammation [51], and enhanced weight in *M.tb*-infected C3HeB/FeJ mice with a sufficient n-3 LCPUFA status [52].

Therapeutic target

Nuclear receptor

In *M.tb* infections, several nuclear receptor agonists have demonstrated encouraging results in enhancing host antimicrobial defense and autophagy. Overexpression of nuclear receptor subfamily 1, group D member 1 (NR1D1)/Rev-Erba, an orphan nuclear receptor, has been shown to enhance the antimycobacterial properties of macrophages and decrease the survival of *M.tb* by promoting phagosome-lysosome maturation through IL-10 repression in human macrophages [53]. Chandra et al. identified NR1D1 as a potential antimicrobial therapeutic target because it promotes autophagy and lysosomal biogenesis through positive regulation of TFEB expression. GSK4112, a synthetic agonist of NR1D1, also induced autophagy in human macrophages [145], supporting the involvement of NR1D1 in the clearance of *M.tb*. Another nuclear receptor, peroxisome proliferator-activated receptor α (PPAR- α), modulates gene expression related to inflammation and mediates antimycobacterial responses against *M.tb* infection by activating the expression of genes associated with autophagy, lysosomal function, and phagosomal maturation through lipid catabolism and upregulation of TFEB transcription [54]. Gemfibrozil (GEM), a PPAR- α activator, has been shown to decrease *Mabc* burden and inflammatory responses in vivo [146]. In addition, the oldest orphan nuclear receptor estrogen-related receptor α (ERR α), an important regulator of metabolic gene transcription and innate immune function, including those induced by toll-like receptor (TLR) and antimicrobial activities against intracellular bacterial infection [147], facilitated post-translational activation of autophagy by deacetylating various autophagy-related proteins, including ATG5, BECN1, and ATG7. Additionally, ERR α is regulated downstream of AMPK and is crucial for antimicrobial host defense against *M.tb* infection by promoting phagosome maturation and regulating excessive inflammation [55]. In summary, strategies targeting ERR α may offer novel therapeutic options for TB treatment.

IL-1 receptor antagonist (IL-1Ra)

IL-1Ra, encoded by *Il1rn*, binds to IL-1R1 without inducing signaling pathways and prevents the binding of IL-1 α/β [148]. An intervention using an anti-IL-1Ra antibody in *M.tb*-infected B6.Sst1^s mice, designed to block IL-1Ra and restore IL-1 signaling, resulted in decreased bacterial burden in the lungs, maintained body weight, and reduced lung lesions in mice [56], suggesting that IL-1Ra might be a promising target for HDT during *M.tb* infection.

Furthermore, certain combination therapies involving HDT have also shown promising results. For instance, everolimus combined with oral L-GSH increased the levels of Th1 cytokines, including IFN- γ , TNF- α , and IL-2 and decreased intracellular *M. bovis* BCG infection in patients with T2DM [149]. The combination of phenylbutyrate (PBA) and 1,25-dihydroxyvitamin D3 (1,25(OH)₂D3) induces LL-37 expression in a lung epithelial cell line and enhances *M.tb* killing in human monocyte-derived macrophages through activation of autophagy and resolution of lung pathology [150]. Clinical trials have also demonstrated that therapy with PBA in combination with vitD3 can promote the clearances of *M.tb* from the respiratory tract, accelerate sputum culture conversion, reduce clinical symptoms, and promote favorable immunomodulation, thus improving treatment outcomes [146]. However, daily supplementation with vitD3 + PBA improved clinical TB symptoms and other complications but did not substantially affect bacterial clearance in sputum [147]. These mixed results underscore the need for further research to optimize HDT strategies.

Discussion

Currently, while traditional anti-tuberculosis therapies, such as the standard first-line regimen (isoniazid, rifampicin, pyrazinamide, and ethambutol), have been effective in treating drug-susceptible TB, they face significant challenges, including drug resistance, adverse effects, and prolonged treatment durations [1]. Consequently, researchers have explored innovative therapeutic approaches, including HDT. HDTs offer a complementary approach by targeting host immune responses to enhance bacterial clearance and reduce tissue damages. Recently, HDT drugs, used as adjuvants to conventional antitubercular regimens, have provided new ideas and hopes, particularly for patients with TB with drug resistance or with various underlying diseases. Findings from a recent study indicate that HDT can reduce the course of therapy, lower the occurrence of drug resistance, and cure refractory tuberculosis. Several HDT drugs used in the treatment of TB, including statins, macrocyclic lactones, and

TKI, have received approval by the US Food and Drug Administration for other clinical applications, underscoring their favorable safety profiles. In addition, while we have categorized HDTs into two broad groups based on their primary effects—(1) decreasing TB burden and (2) reducing tissue damage—it is important to note that some HDTs can simultaneously address both aspects. For example, metformin, a well-studied HDT agent, not only limits excessive inflammation and tissue damage by modulating immune responses but also promotes autophagy through mTOR inhibition, thereby enhancing the clearance of *M. tb* [118]. These dual-function HDTs highlight the potential for multifunctional therapies that target both bacterial survival and host immunopathology. However, for the sake of clarity and intuitive presentation, we have classified HDTs based on their dominant mechanism of action. This approach allows for a more structured discussion while acknowledging the multifaceted nature of some HDTs.

In terms of cost-effectiveness, traditional therapies are generally inexpensive but may incur higher long-term costs due to treatment failures, drug resistance, and management of adverse effects. On the other hand, HDTs, while potentially more expensive initially, could reduce overall healthcare costs by improving treatment efficacy, shortening therapy durations, and minimizing complications. Some HDTs, such as vitamin D, are also a practical and cost-effective solution. However, the widespread adoption of HDTs requires further validation through large-scale clinical trials to establish their efficacy, safety, and cost-effectiveness in diverse populations. By integrating HDTs with existing anti-TB regimens, it may be possible to achieve a more balanced and effective approach to TB management, particularly for drug-resistant and complicated cases.

With respect to safety, it is also crucial to thoroughly evaluate the safety profiles and potential adverse effects of HDTs. For instance, metformin, is generally well-tolerated but can cause gastrointestinal side effects (e.g., nausea, diarrhea) and, in rare cases, lactic acidosis, particularly in patients with renal impairment [118]. Similarly, vitamin D supplementation, although safe at recommended doses, can lead to hypercalcemia and hypercalciuria if administered in excess [151]. Other HDT agents, such as statins, are associated with muscle-related adverse effects (e.g., myopathy and rhabdomyolysis), especially at high doses or when used in combination with other medications metabolized by the cytochrome P450 system [152]. Additionally, immunomodulatory agents like tofacitinib, a JAK inhibitor, carry risks of increased susceptibility to infections, particularly in immunocompromised individuals, as well as potential cardiovascular and thromboembolic events [153].

Therefore, in the future, HDT research should focus on several key areas: (1) refining drug delivery systems to enhance the efficacy of HDT agents in the lungs; (2) conducting larger, multicenter clinical trials to determine optimal dosing and safety profiles; (3) investigating the potential for combination therapies to shorten TB treatment durations while minimizing side effects. Addressing these priorities will be critical for advancing HDT from preclinical promise to clinical reality, and (4) developing more robust and physiologically relevant *in vitro* assays and *in vivo* models that better recapitulate the complex host–pathogen interactions and immune responses observed in human TB.

Acknowledgements

We are grateful to members of our laboratory for helpful discussion and critical comments on the manuscript.

Author contributions

NT, HC, QL, HS, JZ, NC and ZS contributed to the conception and design; NT and HC drafted the initial version of the manuscript, QL, HS, JZ, NC and ZS revised the manuscript critically for intellectual content; All Authors approved the final version of the manuscript; All Authors agree to be accountable for all aspects of the work.

Funding

This work was supported by the National Natural Science Foundation of China (No. 82272347) and the Capital Health Research and Development of Special Fund (No. 2022-1G-2161).

Availability of data and materials

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 30 September 2024 Accepted: 9 March 2025

Published online: 11 April 2025

References

- World Health Organization. Global tuberculosis report 2023. Geneva: World Health Organization; 2023.
- Mitini-Nkhoma SC, Chimbayo ET, Mzinza DT, Mhango DV, Chirambo AP, Mandalasi C, Lakudzala AE, Tembo DL, Jambo KC, Mwandumba HC. Something old, something new: ion channel blockers as potential anti-tuberculosis agents. *Front Immunol*. 2021;12: 665785.
- Frank DJ, Horne DJ, Dutta NK, Shaku MT, Madensein R, Hawn TR, Steyn AJC, Karakousis PC, Kana BD, Meintjes G, et al. Remembering the host in tuberculosis drug development. *J Infect Dis*. 2019;219:1518–24.
- Tobin DM. Host-directed therapies for tuberculosis. *Cold Spring Harb Perspect Med*. 2015;5: a021196.
- Kaufmann SHE, Dorhoi A, Hotchkiss RS, Bartenschlager R. Host-directed therapies for bacterial and viral infections. *Nat Rev Drug Discov*. 2018;17:35–56.
- O'Connor G, Gleeson LE, Fagan-Murphy A, Cryan SA, O'Sullivan MP, Keane J. Sharpening nature's tools for efficient tuberculosis control: a review of the potential role and development of host-directed therapies and strategies for targeted respiratory delivery. *Adv Drug Deliv Rev*. 2016;102:33–54.
- Jolliffe DA, Ganmaa D, Wejse C, Raqib R, Haq MA, Salahuddin N, Daley PK, Ralph AP, Ziegler TR, Martineau AR. Adjunctive vitamin D in tuberculosis treatment: meta-analysis of individual participant data. *Eur Respir J*. 2019;53:1802003.
- Parihar SP, Guler R, Khutlang R, Lang DM, Hurdal R, Mhlanga MM, Suzuki H, Marais AD, Brombacher F. Statin therapy reduces the *Mycobacterium tuberculosis* burden in human macrophages and in mice by enhancing autophagy and phagosome maturation. *J Infect Dis*. 2014;209:754–63.
- Martinet W, Verheye S, De Meyer GR. Everolimus-induced mTOR inhibition selectively depletes macrophages in atherosclerotic plaques by autophagy. *Autophagy*. 2007;3:241–4.
- Stanley SA, Barczak AK, Silvis MR, Luo SS, Sogi K, Vokes M, Bray MA, Carpenter AE, Moore CB, Siddiqi N, et al. Identification of host-targeted small molecules that restrict intracellular *Mycobacterium tuberculosis* growth. *PLoS Pathog*. 2014;10: e1003946.
- Sogi KM, Lien KA, Johnson JR, Krogan NJ, Stanley SA. The tyrosine kinase inhibitor gefitinib restricts *Mycobacterium tuberculosis* growth through increased lysosomal biogenesis and modulation of cytokine signaling. *ACS Infect Dis*. 2017;3:564–74.
- Juárez E, Carranza C, Sánchez G, González M, Chávez J, Sarabia C, Torres M, Sada E. Loperamide restricts intracellular growth of *Mycobacterium tuberculosis* in lung macrophages. *Am J Respir Cell Mol Biol*. 2016;55:837–47.
- Juárez E, Ruiz A, Cortez O, Sada E, Torres M. Antimicrobial and immunomodulatory activity induced by loperamide in mycobacterial infections. *Int Immunopharmacol*. 2018;65:29–36.
- Maiga M, Ahidjo BA, Maiga MC, Cheung L, Pelly S, Lun S, Bougoudogo F, Bishai WR. Efficacy of adjunctive tofacitinib therapy in mouse models of tuberculosis. *EBioMedicine*. 2015;2:868–73.
- Sheikhpour M, Shokrgozar MA, Biglari A, Pornour M, Abdolrahimi F, Poorazar Dizaji S, Khanipour S, Masoumi M, Ebrahimzadeh N, Abolfathi H. Gene expression and *in vitro* pharmacogenetic studies of dopamine and serotonin gene receptors in tuberculosis. *Tanaffos*. 2021;20:126–33.
- Panda S, Tiwari A, Luthra K, Sharma SK, Singh A. Association of Fok1 VDR polymorphism with Vitamin D and its associated molecules in pulmonary tuberculosis patients and their household contacts. *Sci Rep*. 2019;9:15251.
- Zhu X, Zhu Y, Li C, Yu J, Ren D, Qiu S, Nie Y, Yu X, Xu X, Zhu W. 1,25-Dihydroxyvitamin D regulates macrophage polarization and ameliorates experimental inflammatory bowel disease by suppressing miR-125b. *Int Immunopharmacol*. 2019;67:106–18.
- Cohen MS, Mesler DE, Snipes RG, Gray TK. 1,25-Dihydroxyvitamin D3 activates secretion of hydrogen peroxide by human monocytes. *J Immunol*. 1986;136:1049–53.
- Lam KK, Zheng X, Forestieri R, Balgi AD, Nodwell M, Vollett S, Anderson HJ, Andersen RJ, Av-Gay Y, Roberge M. Nitazoxanide stimulates autophagy and inhibits mTORC1 signaling and intracellular proliferation of *Mycobacterium tuberculosis*. *PLoS Pathog*. 2012;8: e1002691.
- Ranjbar S, Haridas V, Nambu A, Jasenosky LD, Sadhukhan S, Ebert TS, Hornung V, Cassell GH, Falvo JV, Goldfeld AE. Cytoplasmic RNA sensor pathways and nitazoxanide broadly inhibit intracellular *Mycobacterium tuberculosis* growth. *iScience*. 2019;22:299–313.
- Gupta A, Meena J, Sharma D, Gupta P, Gupta UD, Kumar S, Sharma S, Panda AK, Misra A. Inhalable particles for “pincer therapeutics” targeting nitazoxanide as bactericidal and host-directed agent to macrophages in a mouse model of tuberculosis. *Mol Pharm*. 2016;13:3247–55.
- Padmapriyadarsini C, Szumowski JD, Akbar N, Shanmugasundaram P, Jain A, Bathragiri M, Pattnaik M, Turuk J, Karunaianantham R, Balakrishnan S, et al. A dose-finding study to guide use of verapamil as an adjunctive therapy in tuberculosis. *Clin Pharmacol Ther*. 2024;115:324–32.
- Kan Y, Meng L, Xie L, Liu L, Dong W, Feng J, Yan Y, Zhao C, Peng G, Wang D, et al. Temporal modulation of host aerobic glycolysis determines the outcome of *Mycobacterium marinum* infection. *Fish Shellfish Immunol*. 2020;96:78–85.

24. Palucci I, Maulucci G, De Maio F, Sali M, Romagnoli A, Petrone L, Fimia GM, Sanguinetti M, Goletti D, De Spirito M, et al. Inhibition of transglutaminase 2 as a potential host-directed therapy against *Mycobacterium tuberculosis*. *Front Immunol*. 2019;10:3042.
25. Wallis RS, Ginindza S, Beattie T, Arjun N, Likoti M, Sebe M, Edward VA, Rassool M, Ahmed K, Fielding K, et al. Lung and blood early biomarkers for host-directed tuberculosis therapies: secondary outcome measures from a randomized controlled trial. *PLoS ONE*. 2022;17: e0252097.
26. To K, Cao R, Yegiazaryan A, Owens J, Nguyen T, Sasaninia K, Vaughn C, Singh M, Truong E, Medina A, et al. Effects of oral liposomal glutathione in altering the immune responses against *Mycobacterium tuberculosis* and the *Mycobacterium bovis* BCG strain in individuals with type 2 diabetes. *Front Cell Infect Microbiol*. 2021;11: 657775.
27. Mitini-Nkhoma SC, Fernando N, Ishaka GKD, Handunnetti SM, Pathirana SL. Ion transport modulators as antimycobacterial agents. *Tuberc Res Treat*. 2020;2020:3767915.
28. Biswas VK, Sen K, Ahad A, Ghosh A, Verma S, Pati R, Prusty S, Nayak SP, Podder S, Kumar D, et al. NCoR1 controls *Mycobacterium tuberculosis* growth in myeloid cells by regulating the AMPK–mTOR–TFEB axis. *PLoS Biol*. 2023;21: e3002231.
29. Madhavan A, Arun KB, Pushparajan AR, Balaji M, Kumar RA. Transcription repressor protein ZBTB25 associates with HDAC1–Sin3a complex in *Mycobacterium tuberculosis*-infected macrophages, and its inhibition clears pathogen by autophagy. *J mSphere*. 2021;6:e00036-e121.
30. Kwon S, Chun HL, Ha HJ, Lee SY, Park HH. Heterogeneous multimeric structure of isocitrate lyase in complex with succinate and itaconate provides novel insights into its inhibitory mechanism. *PLoS ONE*. 2021;16: e0251067.
31. Wentzel AS, Janssen JJE, de Boer VCI, van Veen WG, Forlenza M, Wiegertjes GF. Fish macrophages show distinct metabolic signatures upon polarization. *Front Immunol*. 2020;11:152.
32. Gidon A, Louet C, Røst LM, Bruheim P, Flo TH. The tumor necrosis factor alpha and interleukin 6 auto-paracrine signaling loop controls *Mycobacterium avium* infection via induction of IRF1/IRG1 in human primary macrophages. *MBio*. 2021;12:e0212121.
33. Maiga M, Agarwal N, Ammerman NC, Gupta R, Guo H, Maiga MC, Lun S, Bishai WR. Successful shortening of tuberculosis treatment using adjuvant host-directed therapy with FDA-approved phosphodiesterase inhibitors in the mouse model. *PLoS ONE*. 2012;7: e30749.
34. Maiga M, Ammerman NC, Maiga MC, Tounkara A, Siddiqui S, Polis M, Murphy R, Bishai WR. Adjuvant host-directed therapy with types 3 and 5 but not type 4 phosphodiesterase inhibitors shortens the duration of tuberculosis treatment. *J Infect Dis*. 2013;208:512–9.
35. Subbian S, Tsenova L, Holloway J, Peixoto B, O'Brien P, Dartois V, Khetani V, Zeldis JB, Kaplan G. Adjuvant phosphodiesterase-4 inhibitor therapy improves antibiotic response to pulmonary tuberculosis in a rabbit model. *EBioMedicine*. 2016;4:104–14.
36. Parveen S, Shen J, Lun S, Zhao L, Koleske B, Leone RD, Rais R, Powell JD, Murphy JR, Slusher BS, et al. Glutamine metabolism inhibition has dual immunomodulatory and antibacterial activities against *Mycobacterium tuberculosis*. *Nat Commun*. 2023;14:7427.
37. Datta M, Via LE, Kamoun WS, Liu C, Chen W, Seano G, Weiner DM, Schimel D, England K, Martin JD, et al. Anti-vascular endothelial growth factor treatment normalizes tuberculosis granuloma vasculature and improves small molecule delivery. *Proc Natl Acad Sci U S A*. 2015;112:1827–32.
38. Wang Y, An H, Liu T, Qin C, Sesaki H, Guo S, Radovick S, Hussain M, Maheshwari A, Wondisford FE, et al. Metformin improves mitochondrial respiratory activity through activation of AMPK. *Cell Rep*. 2019;29:1511–1523.e1515.
39. Zheng X, Li W, Xu H, Liu J, Ren L, Yang Y, Li S, Wang J, Ji T, Du G. Sinomenine ester derivative inhibits glioblastoma by inducing mitochondria-dependent apoptosis and autophagy by PI3K/AKT/mTOR and AMPK/mTOR pathway. *Acta Pharm Sin B*. 2021;11:3465–80.
40. Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol*. 2011;13:132–41.
41. Mai NTH, Dobbs N, Phu NH, Colas RA, Thao LTP, Thuong NTT, Nghia HDT, Hanh NHH, Hang NT, Heemskerk AD, et al. A randomised double blind placebo controlled phase 2 trial of adjunctive aspirin for tuberculous meningitis in HIV-uninfected adults. *Elife*. 2018;7: e33478.
42. Byrne ST, Denkin SM, Zhang Y. Aspirin and ibuprofen enhance pyrazinamide treatment of murine tuberculosis. *J Antimicrob Chemother*. 2007;59:313–6.
43. Vilaplana C, Marzo E, Tapia G, Diaz J, Garcia V, Cardona PJ. Ibuprofen therapy resulted in significantly decreased tissue bacillary loads and increased survival in a new murine experimental model of active tuberculosis. *J Infect Dis*. 2013;208:199–202.
44. Tonby K, Wergeland I, Lieske NV, Kvale D, Tasken K, Dyrhol-Riise AM. The COX-inhibitor indomethacin reduces Th1 effector and T regulatory cells in vitro in *Mycobacterium tuberculosis* infection. *BMC Infect Dis*. 2016;16:599.
45. Parasa VR, Muvva JR, Rose JF, Braian C, Brighenti S, Lerm M. Inhibition of tissue matrix metalloproteinases interferes with *Mycobacterium tuberculosis*-induced granuloma formation and reduces bacterial load in a human lung tissue model. *Front Microbiol*. 2017;8:2370.
46. Ong CW, Elkington PT, Brilha S, Ugarte-Gil C, Tome-Esteban MT, Tezera LB, Pabisiak PJ, Moores RC, Sathyamoorthy T, Patel V, et al. Neutrophil-derived MMP-8 drives AMPK-dependent matrix destruction in human pulmonary tuberculosis. *PLoS Pathog*. 2015;11: e1004917.
47. Yang H, Hu J, Chen YJ, Ge B. Role of Sirt1 in innate immune mechanisms against *Mycobacterium tuberculosis* via the inhibition of TAK1 activation. *Arch Biochem Biophys*. 2019;667:49–58.
48. Yang H, Chen J, Chen Y, Jiang Y, Ge B, Hong L. Sirtuin inhibits *M. tuberculosis*-induced apoptosis in macrophage through glycogen synthase kinase-3 β . *Arch Biochem Biophys*. 2020;694:108612.
49. van Toorn R, Solomons RS, Seddon JA, Schoeman JF. Thalidomide use for complicated central nervous system tuberculosis in children: insights from an observational cohort. *Clin Infect Dis*. 2021;72:e136–45.
50. Hayford FEA, Ozturk M, Dolman RC, Blaauw R, Nienaber A, Loots DT, Brombacher F, Smuts CM, Parihar SP, Malan L. Longer-term omega-3 LCPUFA more effective adjunct therapy for tuberculosis than ibuprofen in a C3HeB/FeJ tuberculosis mouse model. *Front Immunol*. 2021;12: 659943.
51. Nienaber A, Baumgartner J, Dolman RC, Ozturk M, Zandberg L, Hayford FEA, Brombacher F, Blaauw R, Parihar SP, Smuts CM, et al. Omega-3 fatty acid and iron supplementation alone, but not in combination, lower inflammation and anemia of infection in *Mycobacterium tuberculosis*-infected mice. *Nutrients*. 2020;12:2897.
52. Nienaber A, Ozturk M, Dolman R, Blaauw R, Zandberg LL, van Rensburg S, Britz M, Hayford FEA, Brombacher F, Loots DT, et al. n-3 long-chain PUFA promote antibacterial and inflammation-resolving effects in *Mycobacterium tuberculosis*-infected C3HeB/FeJ mice, dependent on fatty acid status. *Br J Nutr*. 2022;127:384–97.
53. Chandra V, Mahajan S, Saini A, Dkhar HK, Nanduri R, Raj EB, Kumar A, Gupta P. Human IL10 gene repression by Rev-erba ameliorates *Mycobacterium tuberculosis* clearance. *J Biol Chem*. 2013;288:10692–702.
54. Kim YS, Lee HM, Kim JK, Yang CS, Kim TS, Jung M, Jin HS, Kim S, Jang J, Oh GT, et al. PPAR- α activation mediates innate host defense through induction of TFEB and lipid catabolism. *J Immunol*. 2017;198:3283–95.
55. Kim SY, Yang CS, Lee HM, Kim JK, Kim YS, Kim YR, Kim JS, Kim TS, Yuk JM, Dufour CR, et al. ESRRA (estrogen-related receptor α) is a key coordinator of transcriptional and post-translational activation of autophagy to promote innate host defense. *Autophagy*. 2018;14:152–68.
56. Ji DX, Yamashiro LH, Chen KJ, Mukaida N, Kravnik I, Darwin KH, Vance RE. Type I interferon-driven susceptibility to *Mycobacterium tuberculosis* is mediated by IL-1Ra. *Nat Microbiol*. 2019;4:2128–35.
57. Mizushima N. A brief history of autophagy from cell biology to physiology and disease. *Nat Cell Biol*. 2018;20:521–7.
58. Bruiners N, Dutta NK, Guerrini V, Salamon H, Yamaguchi KD, Karakousis PC, Gennaro ML. The anti-tubercular activity of simvastatin is mediated by cholesterol-driven autophagy via the AMPK–mTORC1–TFEB axis. *J Lipid Res*. 2020;61:1617–28.
59. Guerra-De-Blas PDC, Bobadilla-Del-Valle M, Sada-Ovalle I, Estrada-García I, Torres-González P, López-Saavedra A, Guzmán-Beltrán S, Ponce-de-León A, Sifuentes-Osorio J. Simvastatin enhances the immune response against *Mycobacterium tuberculosis*. *Front Microbiol*. 2019;10:2097.
60. Dutta NK, Bruiners N, Zimmerman MD, Tan S, Dartois V, Gennaro ML, Karakousis PC. Adjunctive Host-directed therapy with statins improves tuberculosis-related outcomes in mice. *J Infect Dis*. 2020;221:1079–87.

61. Adewole OO, Omotoso BA, Ogunsina M, Aminu A, Odeyemi AO, Awopeju OF, Ayoola O, Adedeji T, Sogaolu OM, Adewole TO, et al. Atorvastatin accelerates *Mycobacterium tuberculosis* clearance in pulmonary TB: a randomised phase IIA trial. *Int J Tuberc Lung Dis*. 2023;27:226–8.
62. Cross GB, Sari IP, Kityo C, Lu Q, Pokharkar Y, Moorakonda RB, Thi HN, Do Q, Dalay VB, Gutierrez E, et al. Rosuvastatin adjunctive therapy for rifampicin-susceptible pulmonary tuberculosis: a phase 2b, randomised, open-label, multicentre trial. *Lancet Infect Dis*. 2023;23:847–55.
63. Jimeno A, Rudek MA, Kulesza P, Ma WW, Wheelhouse J, Howard A, Khan Y, Zhao M, Jacene H, Messersmith WA, et al. Pharmacodynamic-guided modified continuous reassessment method-based, dose-finding study of rapamycin in adult patients with solid tumors. *J Clin Oncol*. 2008;26:4172–9.
64. Saxton RA, Sabatini DM. mTOR signaling in growth, metabolism, and disease. *Cell*. 2017;169(2):361–71.
65. Bhatt K, Bhagavathula M, Verma S, Timmins GS, Deretic VP, Ellner JJ, Salgame P. Rapamycin modulates pulmonary pathology in a murine model of *Mycobacterium tuberculosis* infection. *Dis Model Mech*. 2021;14:dmm049018.
66. Ashley D, Hernandez J, Cao R, To K, Yegiazaryan A, Abraham R, Nguyen T, Owens J, Lambros M, Subbian S, et al. Antimycobacterial effects of everolimus in a human granuloma model. *J Clin Med*. 2020;9:2043.
67. Wallis RS, Ginindza S, Beattie T, Arjun N, Likoti M, Edward VA, Rassool M, Ahmed K, Fielding K, Ahdijo BA, et al. Adjunctive host-directed therapies for pulmonary tuberculosis: a prospective, open-label, phase 2, randomised controlled trial. *Lancet Respir Med*. 2021;9:897–908.
68. Napier RJ, Rafi W, Cheruvu M, Powell KR, Zaunbrecher MA, Bornmann W, Salgame P, Shinnick TM, Kalman D. Imatinib-sensitive tyrosine kinases regulate mycobacterial pathogenesis and represent therapeutic targets against tuberculosis. *Cell Host Microbe*. 2011;10:475–85.
69. Kumar MMK, Madhavi K, Mohan T, Nagasree KP, Sangeeta GJ. Novel synthetic analogues of Fluoxetine as potent and selective anti-TB agents. *J Appl Pharm Sci*. 2018;8:107–15.
70. Shankaran D, Singh A, Dawa S, Arumugam P, Gandotra S, Rao V. The antidepressant sertraline provides a novel host directed therapy module for augmenting TB therapy. *Elife*. 2023;12: e64834.
71. Boulware DR, Nalintya E, Rajasingham R, Kirumira P, Naluyima R, Turya F, Namanda S, Rutakingirwa MK, Skipper CP, Nikweri Y, et al. Adjunctive sertraline for asymptomatic cryptococcal antigenemia: a randomized clinical trial. *Med Mycol*. 2020;58:1037–43.
72. Martineau AR, Wilkinson KA, Newton SM, Floto RA, Norman AW, Skolimowska K, Davidson RN, Sørensen OE, Kampmann B, Griffiths CJ, et al. IFN- γ - and TNF-independent vitamin D-inducible human suppression of mycobacteria: the role of cathelicidin LL-37. *J Immunol*. 2007;178:7190–8.
73. Tuckey RC, Cheng CYS, Slominski AT. The serum vitamin D metabolome: what we know and what is still to discover. *J Steroid Biochem Mol Biol*. 2019;186:4–21.
74. Lloyd-Price J, Arze C, Ananthakrishnan AN, Schirmer M, Avila-Pacheco J, Poon TW, Andrews E, Ajami NJ, Bonham KS, Brislawn CJ, et al. Multi-omics of the gut microbial ecosystem in inflammatory bowel diseases. *Nature*. 2019;569:655–62.
75. Druszczynska M, Godkowicz M, Kulesza J, Wawrocki S, Fol M. Cytokine receptors-regulators of antimycobacterial immune response. *Int J Mol Sci*. 2022;23:1112.
76. Wen Y, Li L, Deng Z. Calcitriol supplementation accelerates the recovery of patients with tuberculosis who have vitamin D deficiency: a randomized, single-blind, controlled clinical trial. *BMC Infect Dis*. 2022;22:436.
77. Sinha S, Thukral H, Shareef I, Desai D, Singh BK, Das BK, Dhooria S, Sarin R, Singla R, Meena SK, et al. Prevention of relapse in drug sensitive pulmonary tuberculosis patients with and without vitamin D3 supplementation: a double blinded randomized control clinical trial. *PLoS ONE*. 2023;18: e0272682.
78. Wang J, Xiong K, Wang Q, Zhao S, Liu Y, Ma A. Adjunctive vitamin A and D during pulmonary tuberculosis treatment: a randomized controlled trial with a 2 × 2 factorial design. *Food Funct*. 2020;11:4672–81.
79. Koszalka P, Subbarao K, Baz M. Preclinical and clinical developments for combination treatment of influenza. *PLoS Pathog*. 2022;18: e1010481.
80. Walsh KF, McAulay K, Lee MH, Vilbrun SC, Mathurin L, Jean Francois D, Zimmerman M, Kaya F, Zhang N, Saito K, et al. Early bactericidal activity trial of nitazoxanide for pulmonary tuberculosis. *Antimicrob Agents Chemother*. 2020;64:e01956–e2019.
81. Kim DK, Han D, Bae J, Kim H, Lee S, Kim JS, Jeong YG, Shin J, Park HW. Verapamil-loaded supramolecular hydrogel patch attenuates metabolic dysfunction-associated fatty liver disease via restoration of autophagic clearance of aggregated proteins and inhibition of NLRP3. *Biomater Res*. 2023;27:4.
82. Gupta S, Cohen KA, Winglee K, Maiga M, Diarra B, Bishai WR. Efflux inhibition with verapamil potentiates bedaquiline in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother*. 2014;58(1):574–6.
83. Balganes M, Dinesh N, Sharma S, Kuruppath S, Nair AV, Sharma U. Efflux pumps of *Mycobacterium tuberculosis* play a significant role in antituberculosis activity of potential drug candidates. *Antimicrob Agents Chemother*. 2012;56(5):2643–51.
84. Adams KN, Takaki K, Connolly LE, Wiedenhoft H, Winglee K, Humbert O, Edelstein PH, Cosma CL, Ramakrishnan L. Drug tolerance in replicating mycobacteria mediated by a macrophage-induced efflux mechanism. *Cell*. 2011;145(1):39–53.
85. Parumasivam T, Chan JG, Pang A, Quan DH, Triccas JA, Britton WJ, Chan HK. In vitro evaluation of inhalable verapamil-rifampentine particles for tuberculosis therapy. *Mol Pharm*. 2016;13:979–89.
86. Gupta S, Tyagi S, Almeida DV, Maiga MC, Ammerman NC, Bishai WR. Acceleration of tuberculosis treatment by adjunctive therapy with verapamil as an efflux inhibitor. *Am J Respir Crit Care Med*. 2013;188:600–7.
87. Lemma GL, Wang Z, Hamman MA, Zaheer NA, Gorski JC, Hall SD. The effect of short- and long-term administration of verapamil on the disposition of cytochrome P450 3A and P-glycoprotein substrates. *Clin Pharmacol Ther*. 2006;79:218–30.
88. Huang L, Nazarova EV, Tan S, Liu Y, Russell DG. Growth of *Mycobacterium tuberculosis* in vivo segregates with host macrophage metabolism and ontogeny. *J Exp Med*. 2018;215:1135–52.
89. Xi H, Kurtoglu M, Lampidis TJ. The wonders of 2-deoxy-D-glucose. *IUBMB Life*. 2014;66:110–21.
90. Borrell-Pagès M, Canals JM, Cordelières FP, Parker JA, Pineda JR, Grange G, Bryson EA, Guillemier M, Hirsch E, Hantraye P, et al. Cystamine and cysteamine increase brain levels of BDNF in Huntington disease via HSJ1b and transglutaminase. *J Clin Invest*. 2006;116:1410–24.
91. Jaeger T, Budde H, Flohé L, Menge U, Singh M, Trujillo M, Radi R. Multiple thioredoxin-mediated routes to detoxify hydroperoxides in *Mycobacterium tuberculosis*. *Arch Biochem Biophys*. 2004;423:182–91.
92. Meister A, Anderson ME. Glutathione. *Annu Rev Biochem*. 1983;52:711–60.
93. Venketaraman V, Dayaram YK, Talaue MT, Connell ND. Glutathione and nitrosoglutathione in macrophage defense against *Mycobacterium tuberculosis*. *Infect Immun*. 2005;73:1886–9.
94. Choi SW, Gu Y, Peters RS, Salgame P, Ellner JJ, Timmins GS, Deretic V. Ambroxol induces autophagy and potentiates rifampin antimycobacterial activity. *Antimicrob Agents Chemother*. 2018;62:e01019–e1118.
95. Oppi S, Nusser-Stein S, Blyszczuk P, Wang X, Jomard A, Marzolla V, Yang K, Velagapudi S, Ward LJ, Yuan XM, et al. Macrophage NCOR1 protects from atherosclerosis by repressing a pro-atherogenic PPAR γ signature. *Eur Heart J*. 2020;41:995–1005.
96. Pagán AJ, Ramakrishnan L. The formation and function of granulomas. *Annu Rev Immunol*. 2018;36:639–65.
97. Palmer MV, Kanipe C, Boggiatto PM. The bovine tuberculoid granuloma. *Pathogens*. 2022;11:61.
98. Guirado E, Schlesinger LS. Modeling the *Mycobacterium tuberculosis* granuloma—the critical battlefield in host immunity and disease. *Front Immunol*. 2013;4:98.
99. Russell DG. Who puts the tubercle in tuberculosis? *Nat Rev Microbiol*. 2007;5:39–47.
100. Muñoz-Elías EJ, McKinney JD. *Mycobacterium tuberculosis* isocitrate lyases 1 and 2 are jointly required for in vivo growth and virulence. *Nat Med*. 2005;11:638–44.
101. Nandakumar M, Nathan C, Rhee KY. Isocitrate lyase mediates broad antibiotic tolerance in *Mycobacterium tuberculosis*. *Nat Commun*. 2014;5:4306.
102. Kim YJ, Park EJ, Lee SH, Silwal P, Kim JK, Yang JS, Whang J, Jang J, Kim JM, Jo EK. Dimethyl itaconate is effective in host-directed antimicrobial

- responses against mycobacterial infections through multifaceted innate immune pathways. *Cell Biosci.* 2023;13:49.
103. Zhu X, Guo Y, Liu Z, Yang J, Tang H, Wang Y. Itaconic acid exerts anti-inflammatory and antibacterial effects via promoting pentose phosphate pathway to produce ROS. *Sci Rep.* 2021;11:18173.
 104. Hoffmann E, Machelart A, Belhaouane I, Deboosere N, Pauwels AM, Saint-André JP, Song OR, Jouny S, Poncet A, Marion S, et al. IRG1 controls immunometabolic host response and restricts intracellular *Mycobacterium tuberculosis* infection. 2019; 761551.
 105. Nair S, Huynh JP, Lampropoulou V, Loginicheva E, Esaulova E, Gounder AP, Boon ACM, Schwarzkopf EA, Bradstreet TR, Edelson BT, et al. Irg1 expression in myeloid cells prevents immunopathology during *M. tuberculosis* infection. *J Exp Med.* 2018;215:1035–45.
 106. Serezani CH, Ballinger MN, Aronoff DM, Peters-Golden M. Cyclic AMP: master regulator of innate immune cell function. *Am J Respir Cell Mol Biol.* 2008;39:127–32.
 107. Agarwal N, Lamichhane G, Gupta R, Nolan S, Bishai WR. Cyclic AMP intoxication of macrophages by a *Mycobacterium tuberculosis* adenylate cyclase. *Nature.* 2009;460:98–102.
 108. Subbian S, Tsenova L, O'Brien P, Yang G, Koo MS, Peixoto B, Fallows D, Dartois V, Muller G, Kaplan G. Phosphodiesterase-4 inhibition alters gene expression and improves isoniazid-mediated clearance of *Mycobacterium tuberculosis* in rabbit lungs. *PLoS Pathog.* 2011;7: e1002262.
 109. Maiga MC, Ahidjo BA, Maiga M, Bishai WR. Roflumilast, a type 4 phosphodiesterase inhibitor, shows promising adjunctive, host-directed therapeutic activity in a mouse model of tuberculosis. *Antimicrob Agents Chemother.* 2015;59:7888–90.
 110. Jiang Q, Qiu Y, Kurland IJ, Drlica K, Subbian S, Tyagi S, Shi L. Glutamine is required for M1-like polarization of macrophages in response to *Mycobacterium tuberculosis* infection. *MBio.* 2022;13:e0127422.
 111. Borah K, Beyß M, Theorell A, Wu H, Basu P, Mendum TA, Nöh K, Beste DJV, McFadden J. Intracellular *Mycobacterium tuberculosis* exploits multiple host nitrogen sources during growth in human macrophages. *Cell Rep.* 2019;29:3580–3591. e3584.
 112. Polena H, Boudou F, Tilleul S, Dubois-Colas N, Lecoite C, Rakotosamimanana N, Pelizzola M, Andriamandimby SF, Raharimanga V, Charles P, et al. *Mycobacterium tuberculosis* exploits the formation of new blood vessels for its dissemination. *Sci Rep.* 2016;6:33162.
 113. Oehlers SH, Cronan MR, Scott NR, Thomas MI, Okuda KS, Walton EM, Beerman RW, Crosier PS, Tobin DM. Interception of host angiogenic signalling limits mycobacterial growth. *Nature.* 2015;517:612–5.
 114. Harding JS, Herbath M, Chen Y, Rayasam A, Ritter A, Csoka B, Hasko G, Michael IP, Fabry Z, Nagy A, et al. VEGF-A from granuloma macrophages regulates granulomatous inflammation by a non-angiogenic pathway during mycobacterial infection. *Cell Rep.* 2019;27:2119–2131. e2116.
 115. Drapela S, Ilter D, Gomes AP. Metabolic reprogramming: a bridge between aging and tumorigenesis. *Mol Oncol.* 2022;16:3295–318.
 116. Yang CS, Kim JJ, Lee HM, Jin HS, Lee SH, Park JH, Kim SJ, Kim JM, Han YM, Lee MS, et al. The AMPK-PPARGC1A pathway is required for anti-microbial host defense through activation of autophagy. *Autophagy.* 2014;10:785–802.
 117. Jia J, Bissa B, Brecht L, Allers L, Choi SW, Gu Y, Zbinden M, Burge MR, Timmins G, Hallows K, et al. AMPK, a regulator of metabolism and autophagy, is activated by lysosomal damage via a novel galectin-directed ubiquitin signal transduction system. *Mol Cell.* 2020;77:951–969. e959.
 118. Singhal A, Jie L, Kumar P, Hong GS, Leow MK, Paleja B, Tsenova L, Kurepina N, Chen J, Zolezzi F, et al. Metformin as adjunct antituberculosis therapy. *Sci Transl Med.* 2014;6:263ra159.
 119. Pan SW, Yen YF, Kou YR, Chuang PH, Su VY, Feng JY, Chan YJ, Su WJ. The risk of TB in patients with type 2 diabetes initiating metformin vs sulfonylurea treatment. *Chest.* 2018;153:1347–57.
 120. Yu X, Li L, Xia L, Feng X, Chen F, Cao S, Wei X. Impact of metformin on the risk and treatment outcomes of tuberculosis in diabetics: a systematic review. *BMC Infect Dis.* 2019;19:859.
 121. Degner NR, Wang JY, Golub JE, Karakousis PC. Metformin use reverses the increased mortality associated with diabetes mellitus during tuberculosis treatment. *Clin Infect Dis.* 2018;66:198–205.
 122. Ma Y, Pang Y, Shu W, Liu YH, Ge QP, Du J, Li L, Gao WW. Metformin reduces the relapse rate of tuberculosis patients with diabetes mellitus: experiences from 3-year follow-up. *Eur J Clin Microbiol Infect Dis.* 2018;37:1259–63.
 123. Padmapriyadarsini C, Mamulwar M, Mohan A, Shanmugam P, Gomathy NS, Mane A, Singh UB, Pavankumar N, Kadam A, Kumar H, et al. Randomized trial of metformin with anti-tuberculosis drugs for early sputum conversion in adults with pulmonary tuberculosis. *Clin Infect Dis.* 2022;75:425–34.
 124. Yu SY, Ip MS, Li X, Cheung KS, Ren QW, Wu MZ, Li HL, Wong PF, Tse HF, Yiu KH. Low-dose aspirin and incidence of lung carcinoma in patients with chronic obstructive pulmonary disease in Hong Kong: a cohort study. *PLoS Med.* 2022;19: e1003880.
 125. Derry S, Wiffen PJ, Moore RA. Single dose oral ibuprofen plus caffeine for acute postoperative pain in adults. *Cochrane Database Syst Rev.* 2015;2015:CD011509.
 126. Licican EL, Nguyen V, Sullivan AB, Gronert K. Selective activation of the prostaglandin E2 circuit in chronic injury-induced pathologic angiogenesis. *Invest Ophthalmol Vis Sci.* 2010;51:6311–20.
 127. Rangel Moreno J, Estrada García I, De La Luz García Hernández M, Aguilar Leon D, Marquez R, Hernández Pando R. The role of prostaglandin E2 in the immunopathogenesis of experimental pulmonary tuberculosis. *Immunology.* 2002;106:257–66.
 128. Pellegrini JM, Martin C, Morelli MP, Schander JA, Tateosian NL, Amiano NO, Rolandelli A, Palmero DJ, Levi A, Ciallella L, et al. PGE2 displays immunosuppressive effects during human active tuberculosis. *Sci Rep.* 2021;11:13559.
 129. Mayer-Barber KD, Andrade BB, Oland SD, Amaral EP, Barber DL, Gonzales J, Derrick SC, Shi R, Kumar NP, Wei W, et al. Host-directed therapy of tuberculosis based on interleukin-1 and type I interferon crosstalk. *Nature.* 2014;511:99–103.
 130. Mortensen R, Clemmensen HS, Woodworth JS, Therkelsen ML, Mustafa T, Tonby K, Jenum S, Agger EM, Dyrhol-Riise AM, Andersen P. Cyclooxygenase inhibitors impair CD4 T cell immunity and exacerbate *Mycobacterium tuberculosis* infection in aerosol-challenged mice. *Commun Biol.* 2019;2:288.
 131. Abdel Shaheed C, Beardsley J, Day RO, McLachlan AJ. Immunomodulatory effects of pharmaceutical opioids and antipyretic analgesics: mechanisms and relevance to infection. *Br J Clin Pharmacol.* 2022;88:3114–31.
 132. Tamara L, Kartasasmita CB, Alam A, Gurnida DA. Effects of Vitamin D supplementation on resolution of fever and cough in children with pulmonary tuberculosis: a randomized double-blind controlled trial in Indonesia. *J Glob Health.* 2022;12:04015.
 133. Miow QH, Vallejo AF, Wang Y, Hong JM, Bai C, Teo FS, Wang AD, Loh HR, Tan TZ, Ding Y, et al. Doxycycline host-directed therapy in human pulmonary tuberculosis. *J Clin Invest.* 2021; 131.
 134. Mily A, Rekha RS, Kamal SM, Arifuzzaman AS, Rahim Z, Khan L, Haq MA, Zaman K, Bergman P, Brighenti S, et al. Significant effects of oral phenylbutyrate and vitamin D3 adjunctive therapy in pulmonary tuberculosis: a randomized controlled trial. *PLoS ONE.* 2015;10: e0138340.
 135. Bekele A, Gebreselassie N, Ashenafi S, Kassa E, Aseffa G, Amogne W, Getachew M, Aseffa A, Worku A, Raqib R, et al. Daily adjunctive therapy with vitamin D(3) and phenylbutyrate supports clinical recovery from pulmonary tuberculosis: a randomized controlled trial in Ethiopia. *J Intern Med.* 2018;284:292–306.
 136. Xiang C, Yan HC. Ubiquitin conjugating enzyme E2 C (UBE2C) may play a dual role involved in the progression of thyroid carcinoma. *Cell Death Discov.* 2022;8:130.
 137. Xu Y, Wang L, Zimmerman MD, Chen KY, Huang L, Fu DJ, Kaya F, Rakhilin N, Nazarova EV, Bu P, et al. Matrix metalloproteinase inhibitors enhance the efficacy of frontline drugs against *Mycobacterium tuberculosis*. *PLoS Pathog.* 2018;14: e1006974.
 138. Ordóñez AA, Pokkali S, Kim S, Carr B, Klunk MH, Tong L, Saini V, Chang YS, McKeivitt M, Smith V, et al. Adjunct antibody administration with standard treatment reduces relapse rates in a murine tuberculosis model of necrotic granulomas. *PLoS ONE.* 2018;13: e0197474.
 139. Ordóñez AA, Pokkali S, Sanchez-Bautista J, Klunk MH, Urbanowski ME, Kübler A, Bishai WR, Elkington PT, Jain SK. Matrix metalloproteinase inhibition in a murine model of cavitary tuberculosis paradoxically worsens pathology. *J Infect Dis.* 2019;219:633–6.
 140. Kim YJ, Lee SH, Jeon SM, Silwal P, Seo JY, Hanh BTB, Park JW, Whang J, Lee MJ, Heo JY, et al. Sirtuin 3 is essential for host defense against

- Mycobacterium abscessus infection through regulation of mitochondrial homeostasis. *Virulence*. 2020;11:1225–39.
141. Bhaskar A, Kumar S, Khan MZ, Singh A, Dwivedi VP, Nandicoori VK. Host sirtuin 2 as an immunotherapeutic target against tuberculosis. *Elife*. 2020;9: e55415.
 142. Zhang S, Liu Y, Zhou X, Ou M, Xiao G, Li F, Wang Z, Wang Z, Liu L, Zhang G. Sirtuin 7 regulates nitric oxide production and apoptosis to promote mycobacterial clearance in macrophages. *Front Immunol*. 2021;12: 779235.
 143. Fu LM, Fu-Liu CS. Thalidomide and tuberculosis. *Int J Tuberc Lung Dis*. 2002;6:569–72.
 144. Tsenova L, Sokol K, Freedman VH, Kaplan G. A combination of thalidomide plus antibiotics protects rabbits from mycobacterial meningitis-associated death. *J Infect Dis*. 1998;177:1563–72.
 145. Chandra V, Bhagyaraj E, Nanduri R, Ahuja N, Gupta P. NR1D1 ameliorates *Mycobacterium tuberculosis* clearance through regulation of autophagy. *Autophagy*. 2015;11:1987–97.
 146. Kim YS, Kim JK, Hanh BTB, Kim SY, Kim HJ, Kim YJ, Jeon SM, Park CR, Oh GT, Park JW, et al. The peroxisome proliferator-activated receptor α -agonist gemfibrozil promotes defense against mycobacterium abscessus infections. *Cells*. 2020;9:648.
 147. Yuk JM, Kim TS, Kim SY, Lee HM, Han J, Dufour CR, Kim JK, Jin HS, Yang CS, Park KS, et al. Orphan nuclear receptor ERR α controls macrophage metabolic signaling and A20 expression to negatively regulate TLR-induced inflammation. *Immunity*. 2015;43:80–91.
 148. Dinarello CA. Overview of the IL-1 family in innate inflammation and acquired immunity. *Immunol Rev*. 2018;281:8–27.
 149. To K, Cao R, Yegiazaryan A, Owens J, Sasaninia K, Vaughn C, Singh M, Truong E, Sathananthan A, Venketaraman V. The effects of oral liposomal glutathione and in vitro everolimus in altering the immune responses against *Mycobacterium bovis* BCG strain in individuals with type 2 diabetes. *Biomol Concepts*. 2021;12:16–26.
 150. Coussens AK, Wilkinson RJ, Martineau AR. Phenylbutyrate is bacteriostatic against *Mycobacterium tuberculosis* and regulates the macrophage response to infection, synergistically with 25-hydroxy-vitamin D3. *PLoS Pathog*. 2015;11: e1005007.
 151. Coussens AK, Wilkinson RJ, Hanifa Y, Nikolayevskyy V, Elkington PT, Islam K, Timms PM, Venton TR, Bothamley GH, Packe GE, et al. Vitamin D accelerates resolution of inflammatory responses during tuberculosis treatment. *Proc Natl Acad Sci U S A*. 2012;109:15449–54.
 152. Dutta NK, Bruiners N, Pinn ML, Zimmerman MD, Prideaux B, Dartois V, Gennaro ML, Karakousis PC. Statin adjunctive therapy shortens the duration of TB treatment in mice. *J Antimicrob Chemother*. 2016;71:1570–7.
 153. Atzeni F, Popa CD, Nucera V, Nurmohamed MT. Safety of JAK inhibitors: focus on cardiovascular and thromboembolic events. *Expert Rev Clin Immunol*. 2022;18:233–44.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.